This article was downloaded by: On: 22 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



To cite this Article Xu, XIAO-HUA, Yang, NIAN-YUN, Qian, SHI-HUI, Xie, NING and Duan, JING-AO(2008) 'Dammarane triterpenes from *Ligustrum lucidum*', Journal of Asian Natural Products Research, 10: 1, 33 – 37 To link to this Article: DOI: 10.1080/10286020701273833 URL: http://dx.doi.org/10.1080/10286020701273833

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



Dammarane triterpenes from Ligustrum lucidum

XIAO-HUA XU†‡, NIAN-YUN YANG†‡, SHI-HUI QIAN‡*, NING XIE† and JING-AO DUAN‡

†Department of Phytochemistry, China Pharmaceutical University, Nanjing 210009, China ‡Jiangsu Institute of Traditional Chinese Medicine, Nanjing 210028, China

(Received 28 August 2006; revised 14 December 2006; in final form 22 December 2006)

Two new dammarane triterpenes 3β -acetyl-20S,24R-dammarane-25-ene-24-hydroperoxy-20-ol (1), 20S,24R-dammarane-25-ene-24-hydroperoxy- 3β ,20-diol (2), as well as three known dammarane triterpenes, 3β -acetyl-20S,25-epoxydammarane- 24α -ol (3), 20S,25-epoxydammarane- 3β , 20α -diol (4), 20S-dammarane-23-ene- 3β ,20,25-triol (5) were isolated from the fruits of *Ligustrum lucidum*. Compounds 3-5 were isolated from the fruits of *L. lucidum* for the first time. Their structures were elucidated by spectroscopic methods.

Keywords: Ligusturm lucidum; Oleaceae; Dammarane; Triterpenes

1. Introduction

The fruits of *Ligustrum lucidum* Ait. (Oleaceae) are known as Nuzhenzi which are commonly used for their tonic effects in Chinese medicine [1]. Previous studies had found volatile components, triterpenes, flavonoids, secoiridoid glucosides, and phenolic compounds from this plant [2–4]. In our detailed survey of the fruits of *L. lucidum*, two new dammarane triterpenes, 1 and 2, had been isolated as well as three known compounds 3-5 [5–7] (figure 1). The structures of those compounds were identified by means of one and two dimensional NMR spectroscopic techniques, including HSQC, HMBC, and NOESY.

2. Results and discussion

All compounds (1-5) were obtained from the petroleum ether soluble fraction of 80% ethanol extract from the fruits of *L. lucidum* by column chromatography on silica gel as described in the experiment. Identifications of 3-5 were achieved by comparison with previously reported spectroscopic data.

^{*}Corresponding author. Email: njqsh2005@126.com



Figure 1. The structures of 1-5.

Compound 1 showed a quasi-molecular ion peak $[M + Na]^+$ at m/z 541. HRTOF-MS assigned the molecular formula of $C_{32}H_{54}O_5$. The ¹H NMR spectrum of 1 (table 2) revealed seven methyl groups ($\delta 2 \times 0.85$, 2×0.87 , 0.95, 1.12, 1.75), the ¹³C NMR spectrum of 1 (table 1) showed 32 carbon signals, which typically represent the dammarane skeleton [7], and the carbon signals at $\delta 21.2$, 171.0 revealed the presence of an acetyl carbonyl group.

In the HMQC spectrum, the oxymethine proton (δ 4.48) was attached to a carbon at δ 80.9, suggesting the corresponding carbon was esterified. The large coupling constants (10.5 and 5.5 Hz) of this proton demonstrated the equatorial β -orientation of the acetyl group [5]. Compound **1** was shown to possess a hydroperoxyl group by its positive response to the ferrous thiocyanate reagent [8,9]. The ¹³C NMR signals of **1** were similar to those of compound **3** except

Table 1. 13 C NMR spectral data of compounds 1–5 in CDCl₃ (125 MHz).

Carbon	1	2	3	4	5
1	38.7	39.0	38.7	39.0	38.9
2	23.7	24.9	23.7	24.8	24.8
3	80.9	78.9	80.9	78.9	78.9
4	37.9	39.0	37.9	39.1	39.0
5	55.9	55.9	55.9	55.9	55.9
6	18.1	18.3	18.2	18.3	18.2
7	35.2	35.2	35.7	35.3	35.2
8	40.4	40.4	40.4	40.4	40.4
9	50.6	50.6	50.7	50.8	50.6
10	37.1	37.1	37.1	37.2	37.1
11	21.5	21.5	21.6	21.6	21.5
12	27.4	27.4	27.3	27.4	27.4
13	42.4	42.4	42.9	42.9	42.4
14	50.3	50.3	50.0	50.0	50.3
15	31.2	31.2	31.5	31.5	31.1
16	25.2	25.3	25.7	25.7	24.8
17	49.7	49.7	49.6	49.6	49.9
18	15.4	15.3	15.4	15.4	15.3
19	16.4	16.4	16.4	16.4	16.4
20	75.1	75.1	86.4	86.4	75.1
21	24.7	24.8	23.5	23.5	25.6
22	36.3	36.3	35.2	35.7	43.4
23	24.9	24.9	26.1	26.1	122.3
24	89.7	89.8	83.3	83.3	142.0
25	143.7	143.6	71.4	71.4	71.2
26	114.1	114.2	24.3	24.3	29.7
27	17.5	17.5	27.4	27.4	29.8
28	16.2	28.0	16.3	28.0	28.0
29	27.9	15.5	27.9	15.3	15.5
30	16.4	16.2	16.4	16.2	16.2
CH ₃ CO	21.2		21.3		
$\overline{C}H_{3}CO$	171.0		170.9		

34

for the signals due to the side-chain part (C-21–C-27). The structure of the side-chain part were determined by HMQC, HMBC, and NOE experiments. A singlet of methylene protons at δ 5.02 (2H, s) suggested the presence of methylene moiety in the side chain. The carbon signal at δ 89.7 was assigned to C-24, because H-24 was observed as triplet-like signal (J = 12.6 Hz) due to coupling with H-23 [9], in the HMBC spectrum, the methine proton signal at δ 4.28 (1H, t, J = 12.6 Hz) showed correlations with C-22 (δ 36.3), C-23 (δ 24.9), C-25 (δ 143.7), C-26 (δ 114.1) and C-27 (δ 17.5), so this proton signal was assigned to H-24. These findings proved the hydroperoxyl group at C-24. Furthermore other correlation were also observed from the following protons and carbons: H-21 (δ 1.12)/C-20, C-22; H-27 (δ 1.75)/C-24, C-25, C-26; H-22 (δ 5.02)/C-25, C-24, C-27 (figure 2), confirmed the structure of the side chain in **1**.

The relative stereochemistry of four-ring systems was clarified by the NOESY spectrum. The configuration of C-17 was assigned as S by the correlation between H-17 (δ 1.74) and H-30 (δ 0.87) in NOESY spectrum. Correlations between H-21 and H-16, 17 were not detected; however, strong interactions between the H-21 signal at δ 1.12 and H-22, H-23 were observed in NOESY spectrum as shown in figure 2, which led to the assignment of C-20 configuration as S [10,11]. Correlations between H-24 at δ 4.28 and H-22, H-23, H-26, and H-27 were observed in NOESY spectrum as shown in figure 2, which led to the assignment of C-24 configuration as R. Thus the structure of 1 was determined as 3 β -acetyl-20*S*,24*R*-dammarane-25-ene-24-hydroperoxy-20-ol.

Compound **2** showed a quasi-molecular ion peak $[M + Na]^+$ at m/z 499. HRTOF-MS assigned the molecular formula $C_{30}H_{52}O_4$. Compared with **1**, the ¹H NMR and ¹³C NMR spectra of compound **2** (tables 1 and 2) were almost identical to those of **1**, except for the absence of an acetyl group in **2**. In ¹H NMR spectrum, signal of H-3 (δ 3.20, dd, J = 10.5, 5.5 Hz) shifted towards upfield, compared with that of **1** (δ 4.48, dd, J = 10.5, 5.5 Hz), suggesting the corresponding carbon was not esterified. In the HMBC spectrum, H-3 (δ 3.20) correlated with C-1 (δ 39.0), C-2 (δ 24.9), C-4 (δ 39.0), C-28 (δ 28.0), C-29 (δ 15.5). Thus the structure of **2** was deduced as 20*S*,24*R*-dammarane-25-ene-24-hydroperoxy-3 β ,20-diol.

3. Experimental

3.1 General experimental procedures

Melting points were determined on an XT4A micromelting apparatus and are uncorrected. IR spectra were measured on a Perkin–Elmer 8900 FT-IR instrument as KBr disks. 1D and 2D NMR spectra were recorded in CDCl₃ on a Varian-500 spectrometer. HRTOF-MS and SCI-MS



Figure 2. Key HMBC and NOESY correlations of 1.

X.-H. Xu et al.

Н	1 J (Hz)	2 J (Hz)	3 J (Hz)	4 J (Hz)	5 J (Hz)
3	4.48 dd 10.5, 5.5	3.20 dd 10.5, 5.5	4.47 dd 10.5, 5.5	3.19 dd 10.5 5.5	3.19 dd 10.5, 5.5
18	0.95 s	0.96 s	0.95 s	0.96 s	0.96 s
19	0.85 s	0.85 s	0.84 s	0.84 s	0.86 s
21	1.12 s	1.11 s	1.05 m	1.08 m	1.12 s
22	1.48 m	1.47 m	1.57 m	1.58 m	2.19 dd 7.0, 5.0
	1.53 m	1.53 m	1.68 m	1.67 m	
23	1.47 m	1.46 m	1.74 m	1.73 m	5.70 m
	1.71 m	1.70 m	1.82 m	1.83 m	
24	4.28 t 12.6	4.28 t 12.6	3.72 t 14.4	3.73 t 14.4	5.70 m
26	5.02 s	5.02 s	1.11 s	1.12 s	1.32 s
27	1.75 s	1.73 s	1.12 s	1.13 s	1.32 s
28	0.87 s	0.99 s	0.86 s	0.96 s	0.97 s
29	0.85 s	0.77 s	0.84 s	0.77 s	0.77 s
30	0.87 s	0.87 s	0.86 s	0.86 s	0.84 s
CO	2.01 s		2.04 s		
OOH	8.11 s	8.12 s			

Table 2. ¹H NMR spectral data of compounds 1-5 in CDCl₃ (500 MHz).

spectra were performed on Bruker APEX- α mass spectrometer and Micromass Qauttro micro mass spectrometer respectively. Column chromatography was carried out on silica gel (200–300 mesh Qingdao Haiyang Chemical Co. Ltd, China) and Sephadex LH-20 (Pharmacia Biotech).

3.2 Plant material

The fruits of *Ligustrum lucidum* were collected from Nanjing city, Jiangsu Province, China in November 2004 and identified by Researcher Qian. A voucher specimen (No. S-07-00020) is deposited in the Jiangsu Institute of Traditional Chinese Medicine.

3.3 Extraction and isolation

Air-dried fruits (8 kg) were extracted with 80% ethanol (2 × 50 L) for 2 h under reflux, and the combined extracts were concentrated *in vacuo*. The obtained extract (1.835 kg) was then suspended in H₂O and extracted successively with petroleum ether, EtOAc, *n*-butanol saturated with H₂O to give the respective extracts after solvent removal. The petroleum ether soluble portion (231 g) was eluted with petroleum ether/EtOAc by column chromatography on silica gel. Based on TLC characteristics, six major fractions I–VI were made. Fraction α (19.4 g) was further isolated by repeated column chromatography on silica gel with petroleum ether/EtOAc (10:1) to afford **3** (56 mg) and the following four subfractions α a–d. Subfraction α b was purified by silica gel with petroleum ether/EtOAc (96:4) as eluent, and afforded 1 (30 mg). Fraction β (13.1 g) was fractionated by repeated column chromatography on silica gel with petroleum ether/EtOAc (96:4) to give four subfractions β a–d. Compounds **2** (13 mg) and **4** (20 mg) were obtained from subfractions β b and β c, respectively, with petroleum ether/EtOAc (93:7). Fraction β d was purified by Sephadex LH-20 with CHCl₃/MeOH (1:1), and **5** (6 mg) was obtained.

3.3.1 3β-Acetoxyl-20*S*,24*R*-dammarane-25-ene-24-hydroperoxy-20-ol (1). White powder, mp 220–222°C. HRTOF-MS *m/z* 541.3886 [M + Na]⁺ (calcd for C₃₂H ₅₄O₅Na, 541.3881); IR (KBr) (ν_{max} cm⁻¹): 3420, 1718, 1455, 1381, 1255, 1123, 1020, 900; ¹H NMR and ¹³C NMR (CDCl₃) spectral data: see tables 1 and 2, respectively.

3.3.2 20S,24R-Dammarane-25-ene-24-hydroperoxy-3\beta,20-diol (2). White powder, mp 197–200°C. HRTOF-MS *m/z* 499.3742 [M + Na]⁺ (calcd for C₃₀H₅₂O₄Na, 499.3736); IR (KBr) (ν_{max} cm⁻¹): 3425, 1453, 1378, 1249, 1126, 1017, 920; ¹H NMR and ¹³C NMR (CDCl₃) spectral data: see tables 1 and 2, respectively.

3.3.3 3β-Acetoxyl-20S,25-epoxydammarane-24 α -ol (3). White powder, mp 265–266°C. ESI-MS *m*/*z* 526.4 [M + Na + H]⁺; IR (KBr) (ν_{max} cm⁻¹): 3472, 1723, 1449, 1383, 1261, 1123, 1020, 960; ¹H NMR and ¹³C NMR (CDCl₃) spectral data: see tables 1 and 2, respectively.

3.3.4 20S,25-Epoxydammarane-3 β ,24 α -diol (4). White powder, mp 196–198°C. ESI-MS m/z 484.1 [M + Na + H]⁺; IR (KBr) (ν_{max} cm⁻¹): 3467, 1451, 1377, 1246, 1119, 1022, 945; ¹H NMR and ¹³C NMR (CDCl₃) spectral data: see tables 1 and 2, respectively.

3.3.5 20S-Dammarane-23-ene-3 β **,20,25-triol (5)**. White powder, mp 107–109°C. ESI-MS *m*/*z* 484.2 [M + Na + H]⁺; IR (KBr) (ν_{max} cm⁻¹): 3467, 1461, 1382, 1249, 1131, 1028, 936; ¹H NMR and ¹³C NMR (CDCl₃) spectral data: see tables 1 and 2, respectively.

3.4 Validation of the hydroperoxyl group

A solution of 0.5 mg of compound 1 in 1 ml anhydrous Et_2O was prepared, and 0.2 ml 0.1 mol/L fresh FeSO₄ solution was added, and acutely surged. The colour of solution changed from green to red.

Acknowledgements

The programme was supported by the Commonweal Foundation of Jiangsu Province (grant No: BM2004525).

References

- [1] Pharmacopoeia Committee of China, *Chinese Pharmacopoeia (Part A)*, pp. 31–32, The Technology Publishing House, Beijing (2005).
- [2] L.J. Wu, T. Xiang, B.L. Hou, S. Yin, X.C. Zhou. Acta Botan. Sin., 40, 83 (1998).
- [3] T.H. Xiong. J. HuNan Teachers College, 21, 59 (1999).
- [4] Z.D. He, H. Dong, H.X. Xu, W.C. Ye, H.D. Sun, P.P.H. But. Phytochemistry, 56, 327 (2001).
- [5] A.S. Shamsur Rouf, Y. Ozaki, M.A. Rashid, J. Rui. Phytochemistry, 56, 815 (2001).
- [6] D. Butruille, X.A. Dominguez. Tedrahedron Lett., 8, 639 (1974).
- [7] P.G. Waterman, S. Ampofo. *Phytochemistry*, **24**, 2925 (1985).
- [8] M. Yoshikawa, T. Murakami, T. Ueno, N. Hirokawa, K. Yashiro, N. Murakami, J. Yam-ahara, H. Matsuda, R. Saijoh, O. Tanaka. *Chem. Pharm. Bull.*, 45, 1056 (1997).
- [9] Q.Y. Xing, R.Q. Xu, Z. Zhou, W.W. Bei. *Basic Organic Chemistry (Part Two)*, Altitude Publishing House, Beijing (1993).
- [10] B.M. Kwon, S.H. Lee, K.S. Kim, I.R. Lee, U.C. Lee, S.H. Hong, S.H. Bok. Biol. Med. Chem. Lett., 7, 971 (1997).
- [11] H. Kizu, M. koshijima, T. Hayashi. Chem. Pharm. Bull., 33, 1400 (1985).